

Introduction



Flax is an important crop for food and textiles

5 major fungi are pathogens of flax :
Alternaria linicola, *Botrytis cinerea*,
Colletotrichum linicola, *Boeremia exigua* and *Fusarium* spp

→ Transmitted by seeds

To limit the spread of these fungi :
regulated as quarantine or as regulated non quarantine pest in several countries



European Union : maximum permitted infection rate at 5% of the seeds.

A detection method on seeds is essential to limit the spread of these pathogens



ISTA rule 7-007

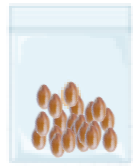
Alternaria linicola
Botrytis cinerea
Colletotrichum linicola

Aim: addition of 2 pathogens in the ISTA method

→ To modify the ISTA method: it is necessary to validate modification by evaluating performance criteria of the method

Method proposed

→ Similar to the current ISTA 7-007 method: morphological identification of fungi



Seed sample

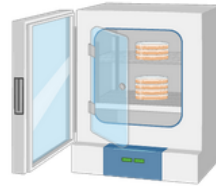
Sample size:
400 seeds



Plating seeds on media

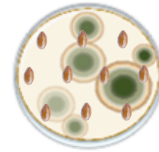
Malt-Agar (MA)
or
Potatoes Dextrose Agar (PDA)
with streptomycin 50 mg/L

10 seeds per plate



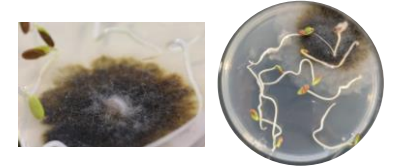
Incubation

7 days at 20°C
in darkness
or
12h dark/12h NUV



Morphological identification

Record the number of infected
seeds for each pathogen
Botrytis cinerea, *Alternaria
linicola*, *Colletotrichum linicola*,
Boeremia exigua, *Fusarium spp.*



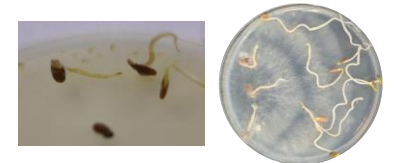
Boeremia exigua



Colletotrichum linicola



Fusarium spp.



Botrytis cinerea



Alternaria linicola

Evaluation of the performance criteria

Criteria	Definition
Analytical specificity	The ability of a method to detect target strains (inclusivity) while not detecting non-targets strains (exclusivity)
Analytical sensitivity (Limit of detection)	Smallest amount of the target pathogen that can be detected
Diagnostic sensitivity	The ability of a method to give correct positive results and not produce false negatives
Diagnostic specificity	The ability of a method to give correct negative results and not produce false positives
Repeatability	The ability of a method to produce the same results when repeated on identical samples under the same conditions (equipment, laboratory, time, etc.).
Reproducibility	The ability of a method to produce the same results when repeated on identical samples under different conditions (equipment, laboratories, time, etc.).

Evaluation of the analytical specificity

Criteria	Definition	Way to evaluate
Analytical specificity	The ability of a method to detect target strains (inclusivity) while not detecting non-targets strains (exclusivity)	Morphological criteria of a set of target and non-target strains were compared with the expected morphological criteria

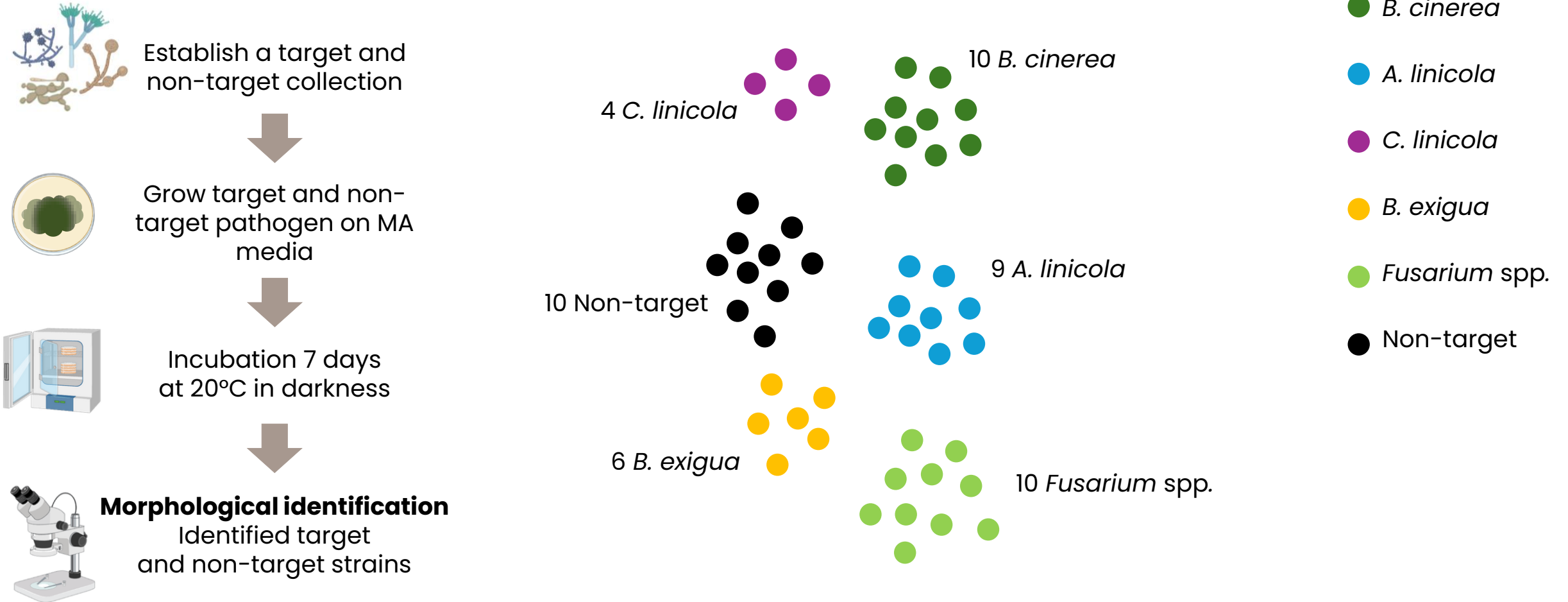
Target collection - 39

Species	Quantity of strains tested	Isolation host	Collection date
<i>Alternaria linicola</i>	9	Flax seed	1999 - 2025
<i>Colletotrichum linicola</i>	4	Flax seed	1999-2024
<i>Botrytis cinerea</i>	10	Flax seed, radish, Faba bean, Chickpea, Cabbage	2010-2025
<i>Boeremia exigua</i>	6	Flax seed and Lettuce	2014-2024
<i>Fusarium avenaceum</i>	2	Flax seed	2024-2025
<i>Fusarium poae</i>	2	Flax seed	2018-2024
<i>Fusarium sporotrichioides</i>	1	Unknown	2019
<i>Fusarium graminearum</i>	3	Flax seed and wheat	2002-2025
<i>Fusarium culmorum</i>	1	Flax seed	2024
<i>Fusarium cerealis</i>	1	Flax seed	2024

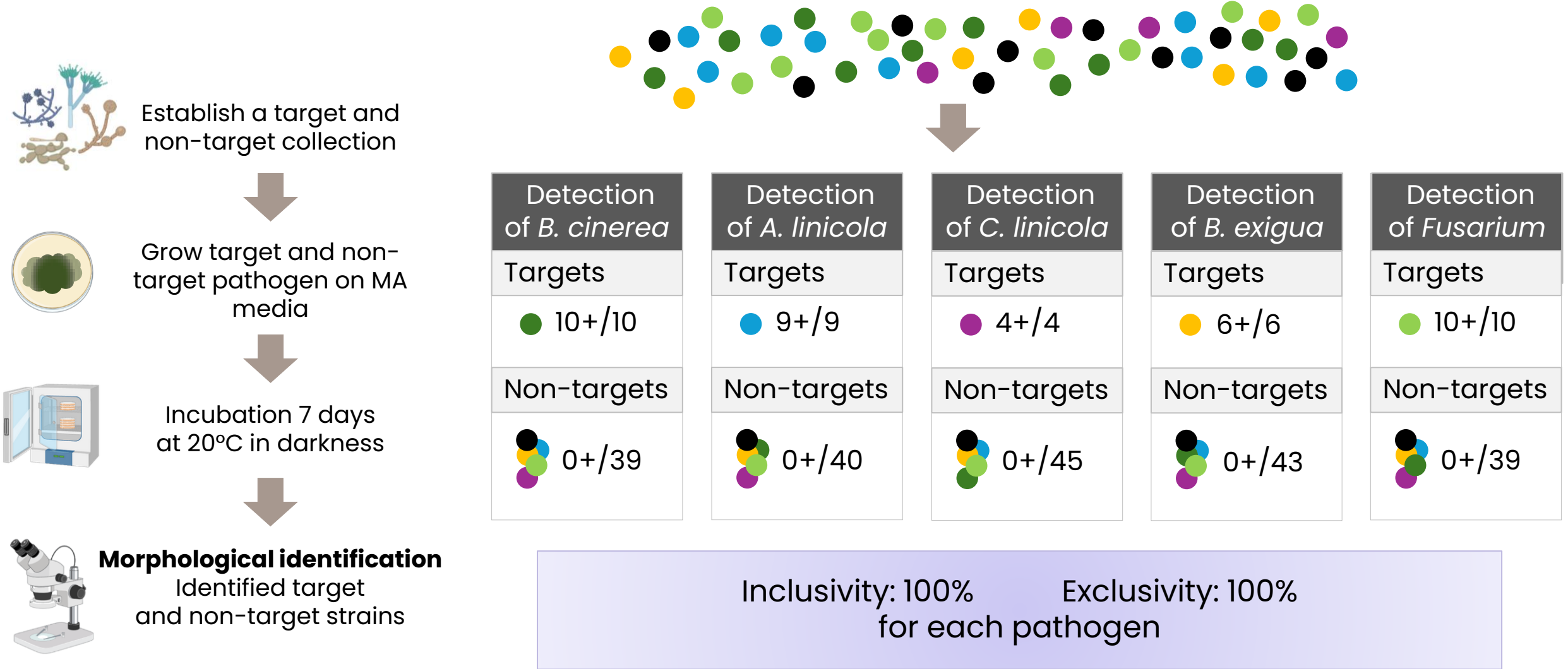
Non-target collection - 10

Species	Isolation host	Collection date
<i>Phoma valerianellae</i>	Lettuce	2020
<i>Alternaria dauci</i>	Carrot	1997
<i>Epicoccum nigrum</i>	NA	2011
<i>Colletotrichum capsici</i>	NA	2019
<i>Trichoderma trixiae</i>	Flax seed	2022
<i>Alternaria alternata</i>	NA	2011
<i>Colletotrichum graminicola</i>	Corn	2009
<i>Rhizoctonia sp.</i>	Flax seed	2021
<i>Alternaria brassicicola</i>	Cabbage	2000
<i>Colletotrichum lupini</i>	Lupin	2023

Evaluation of the analytical specificity



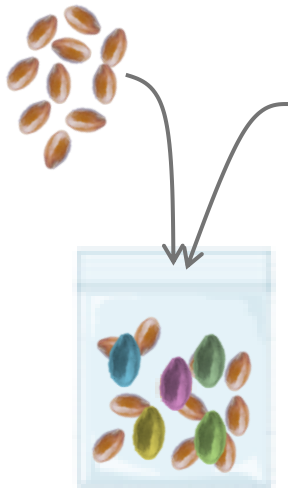
Evaluation of the analytical specificity



Evaluation of the analytical sensitivity

Criteria	Definition	Way to evaluate
Analytical sensitivity (Limit of detection)	Smallest amount of the target pathogen that can be detected	10 samples will be tested, each sample contained 1 contaminated seed for each pathogen and 395 healthy seeds

395
healthy
seeds



- 1 seed contaminated by *B. cinerea*
- 1 seed contaminated by *A. linicola*
- 1 seed contaminated by *C. linicola*
- 1 seed contaminated by *B. exigua*
- 1 seed contaminated by *Fusarium* sp.

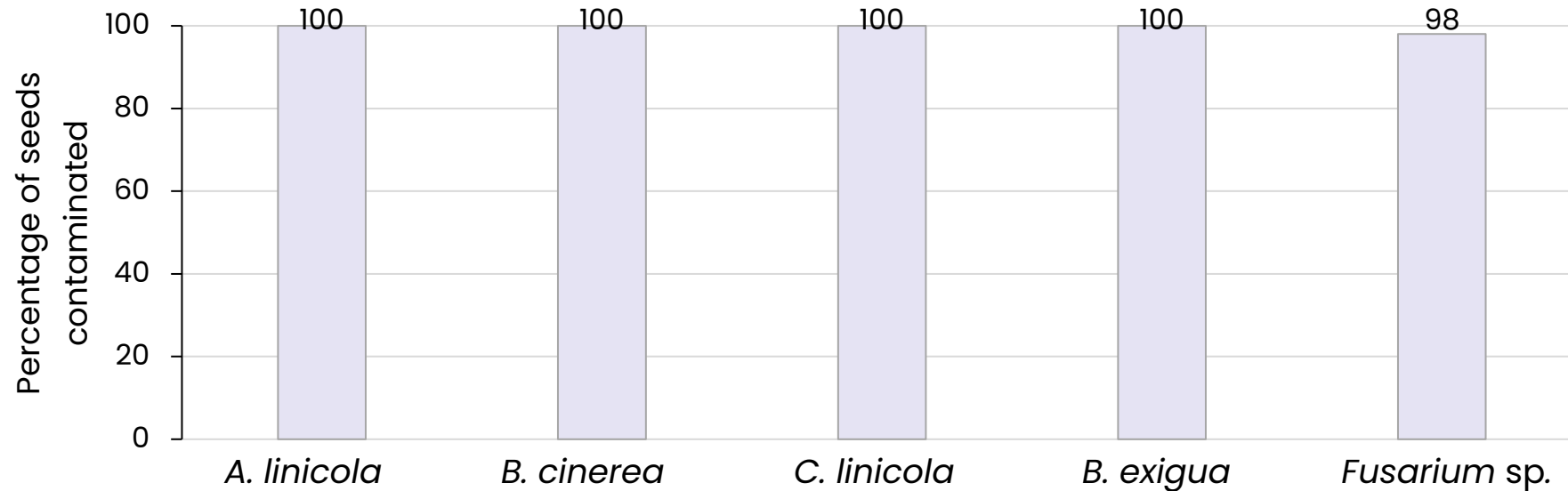
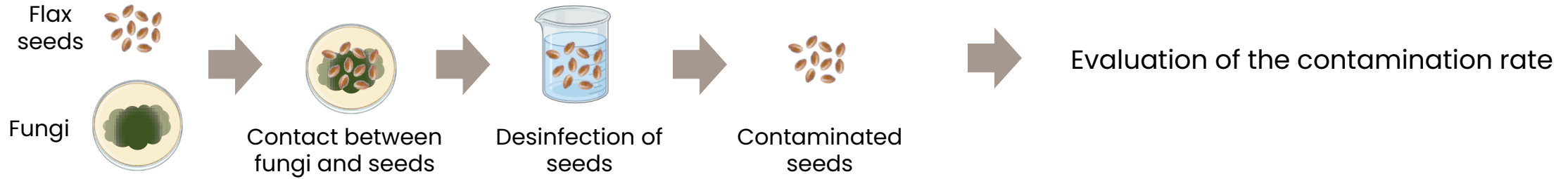


Need to produce artificial
contaminated seeds

Evaluation of the analytical sensitivity

Development of a method for artificial contamination

Machado *et al.*, 2004, Sousa *et al.*, 2006

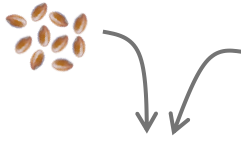


Contamination rates are at 100-98%

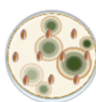
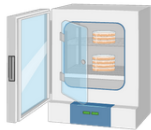
These seeds can be used for the evaluation of the analytical sensitivity

Evaluation of the analytical sensitivity

395
healthy
seeds



- 1 seed contaminated by *B. cinerea*
- 1 seed contaminated by *A. linicola*
- 1 seed contaminated by *C. linicola*
- 1 seed contaminated by *B. exigua*
- 1 seed contaminated by *Fusarium* spp.



Plating seeds on media
Malt-Agar (MA)
with streptomycin 50 mg/L
10 seeds per plate

Incubation
7 days at 20°C in darkness

Morphological identification
Record the number of seeds infected
by *Botrytis cinerea*, *Alternaria linicola*,
Colletotrichum linicola, *Boeremia
exigua*, *Fusarium* spp.



Test performed on 10 samples

Samples	% of seeds contaminated by				
	<i>A. linicola</i>	<i>C. linicola</i>	<i>B. cinerea</i>	<i>B. exigua</i>	<i>Fusarium</i> spp.
1	0.25	0.25	0.25	0.25	0.25
2	0.25	0.25	0.25	0.25	0.25
3	0.25	0.25	0.25	0.25	0.25
4	0.25	0.25	0.25	0.25	0.25
5	0.25	0.25	0.25	0.25	0.25
6	0.25	0.25	0.25	0.25	0.25
7	0.25	0.25	0.25	0.25	0.25
8	0.25	0.25	0.25	0.25	0.25
9	0.25	0.25	0.25	0.25	0.25
10	0.25	0.25	0.25	0.25	0.25

All contaminated seeds were detected

Analytical sensitivity is validated at 1 contaminated seed in a sample of 400 seeds (corresponding to 0.25% of contamination)

Evaluation of the performance criteria

Criteria	Definition	Way to evaluate
Diagnostic sensitivity	The ability of a method to give correct positive results and not produce false negatives	Validated through a TPS with at least 3 laboratories (ideally 8) Samples with 3 levels of infection (healthy, medium and highly infected)
Diagnostic specificity	The ability of a method to give correct negative results and not produce false positives	
Repeatability	The ability of a method to produce the same results when repeated on identical samples under the same conditions (equipment, laboratory, time, etc.).	
Reproducibility	The ability of a method to produce the same results when repeated on identical samples under different conditions (equipment, laboratories, time, etc.).	

Organization of a test performance study



Several qualified laboratories



Panel of codified samples
Infected and healthy

Evaluation of the performance criteria



Organized with ISTA

Selection of
qualified
laboratories

Organization of a pre comparative test

Preliminary test to evaluate stability of the artificial contamination and laboratories expertise on fungi for the method

11 participants

Panel of 9 samples of 100 seeds

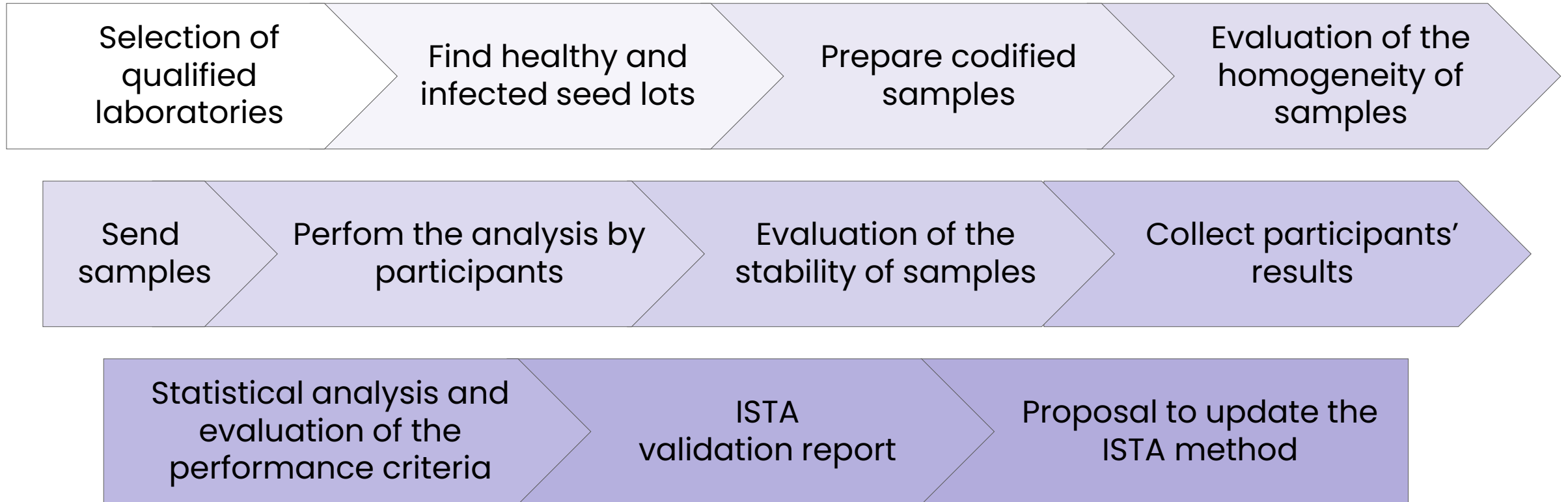
Healthy and artificially contaminated samples

Results of the different laboratories will be compared using boxplot, and statistical test will be used.

Evaluation of the performance criteria



Organized with ISTA



Conclusion



An ISTA method for five-fungi detection in flax seeds:
validation in progress.

Quantitative method
Ensure the viability of fungi

But...

need of expertise in fungi identification by morphology

Validated internationally today,
reliable worldwide tomorrow.



A practical tool to improve global plant health
and support regulatory control.



Thank you for your attention



Claire
Granon



Céline
Le Guisquet



Gaël
Cesbron



Lorine
Le Daré



Isabelle
Sérandat



Justine
Foucher



Nicolas
Denancé



Jaiana
Malabarba



Mylène
Ruh



Quentin
Beaupere

