# COMPARISON OF DETECTION METHODS FOR DITYLENCHUS IN ALFALFA AND FABA BEAN SEED LOTS, AND METHOD VALIDATION

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#### INTRODUCTION

The stem and bulb nematode, Ditylenchus dipsaci, causes swelling and deformation of aerial parts and necrosis or rot of stem base. A race, on 30 biological race described, distinguished from D. dipsaci, owing to a greater body size was named "giant race" and described as a new species named D. gigas (Volvas et al, 2011). The detection and identification of D. dipsaci and D. gigas in seed lots is an







obligatory part of the sanitary control and regulation in Europe (import, export and sale of seed lots to farmers) on alfalfa seeds. Few studies have reported the difference on morphological characters between the two pathogens. However molecular methods have been recently developed to confirm the *Ditylenchus* subsp. (Esquibet et al, 2003; Kerkoud et al, 2007; Volvas et al, 2011). The aim of the TESTA project was to harmonize and validate at international level a detection method of D. dipsaci and D. gigas.

**GOAL** 

**Seed lots** 

Alfaalfa 1

Alfaalfa 2

Alfaalfa 3

Faba bean

Faba bean 2

Faba bean 3

- To compare performance of the biological and molecular protocols currently used in Europe.
- To validate a method which enables the detection of D. dipsaci and D.gigas and propose it as an official ISTA and EPPO protocol.

### COMPARISON OF PROTOCOLS

#### Contamination rate in seed lots **Decantation protocol Decantation protocol Filtration protocol** (Sieving) (bottom) (Supernatant) Nb positive/ Nb positive/ Nb positive/ Couting Estimation Estimation nb samples nb samples nb samples 5/5 843 5/5 50 to 500 5/5 1 à 15 5/5 1204 5/5 > 500 5/5 50 à 500 5/5 5/5 50 à 500 1404 >500 5/5 5/5 5/5 10190 > 500 > 500

4/5

50 to 500

15 to 50

Filtration protocol = concentrate population of nematode High capacity of detection in low infected seed lots

454

124

4/5

5/5

**Decantation protocol** = discarded supernatent Detection of *Ditylenchus* sp. in supernatant False negative sample

5/5

Filtration protocol vs Decantation protocol

15 to 50

15 to 50

# **Detection limit** Filtration protocol (rep1) Filtration protocol (rep2) Decantation protocol (rep1) 6 No postive sample / 5 samples Decantation protocol (rep2) 5 5 5 No. of nematodes articially added by samples Filtration protocol limit of detection at 10 nematodes (higher for decantation protocol) Method chosen

#### **Confirmation by PCR** Primers available Accuracy and repeatability calculated for the four evaluated methods Esquibet NIAB ■ Clear Detection Kerkoud 100% 80% 40% 20% sensitivity specificity sensitivity specificity accuracy repeatability D. gigas D. dipsaci overall results Kerkoud Wood primers primers Identification of both Best results

species in mixed samples.

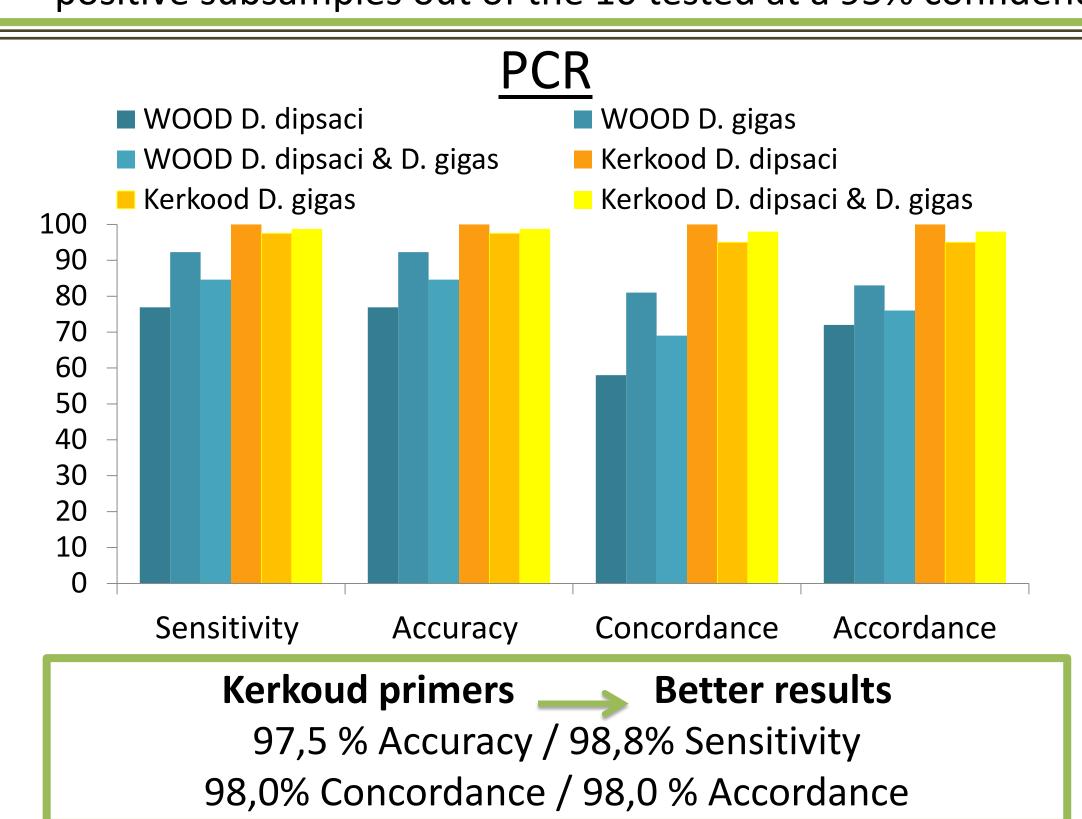
# **VALIDATION TESTS**

Three seed lots (healthy, medium and high infected) of alfalfa and Faba bean were tested for homogeneitiy and confirmed by qualititive analysis. 8 participants from France, Germany, United Kingdom, Czech Republic and Slovenia tested the filtration method and the Kerkoud and Wood primers. Performance criteria were studied: Diagnostic specificity, Diagnostic sensitivity, Accuracy, Accordance (repeatability), Concordance (reproducibility)

# Homogeneity & Stability test 10 samples were tested per seed lot

		Homogeneity test	Stability test
Alfalfa	Lot A	0+/10	0+/10
	Lot B	10+/10	10+/10
	Lot C	10+/10	10+/10
Faba bean	Lot D	0+/10	0/10
	Lot E	9+/10	9+/10
	Lot F	8+/10	10+/10

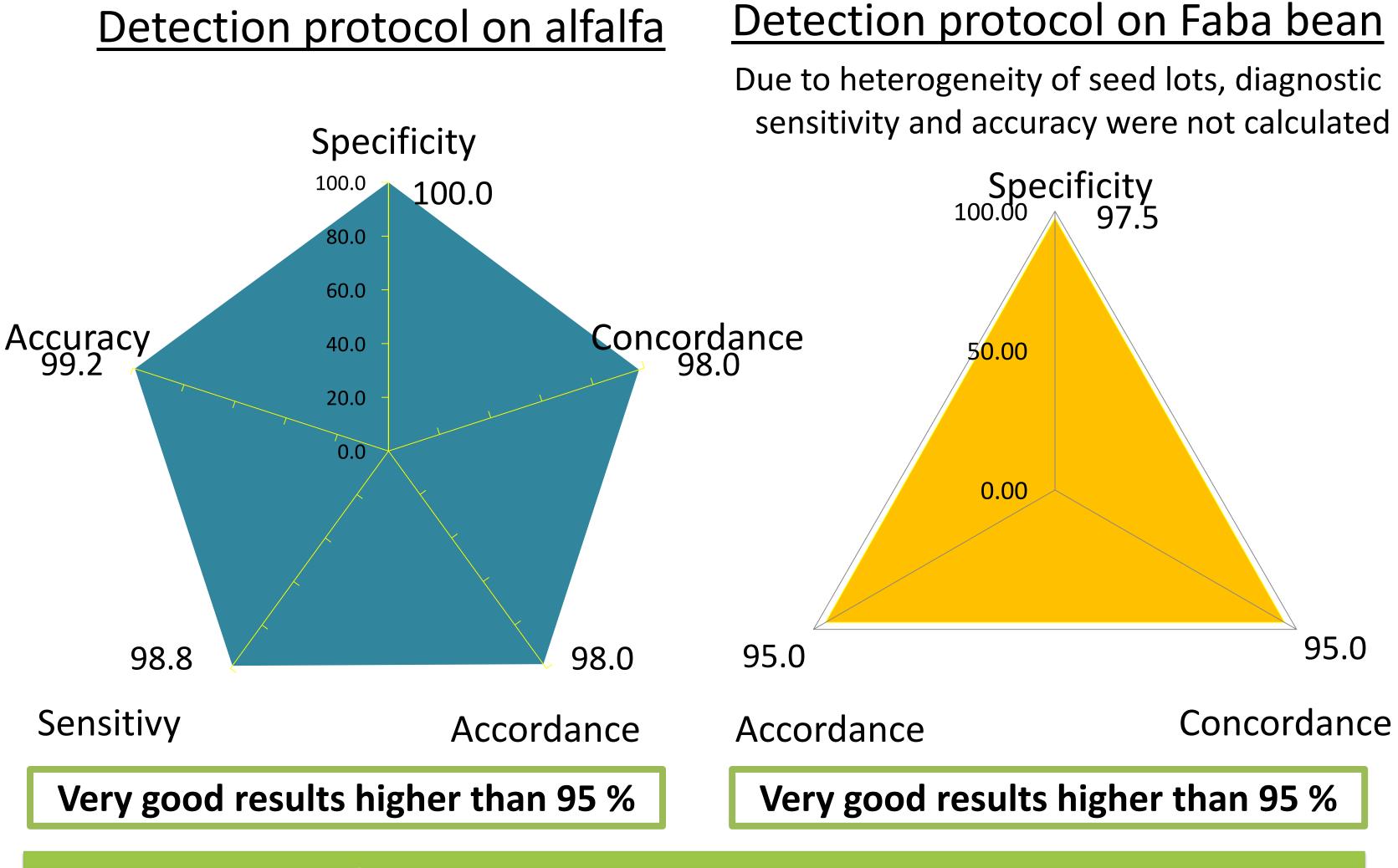
## Conform to expected results. For heterogenous seed lot E and F No. of positive subsample conformed to expected positive subsamples out of the 10 tested at a 95% confidence



Results of the validation test

89 % accuracy

Results obtained by participants conform to expected results



Combination of all these results proved that a detection method using filtration in alfalfa and faba bean seed lot was validated.

CONCLUSION

A detection method using sieving at 20µm to concentrate population of nematodes present in a alfalfa and faba bean seed lot is validated. A PCR method using Kerkoud primers is validated in order to confirm the species of Ditylenchus between D. dipsaci and D. gigas.



